

agnosis is acceptable for treatment assignment purposes and to provide epidemiological indicators for allowing type-specific trend analysis?

The Brazilian Leprosy Control Programme already has recognized the bias reported in the allocation of patients and has been recommending the use of bacilloscopy, whenever possible, to assign patients to MB or PB groups. Indeterminate cases previously allocated as MB are considered PB regardless of the Mitsuda test.

A health system and epidemiology research are required in order to understand regional differences and improve diagnosis accuracy, compliance and coverage. These are essential tools to achieve community effectiveness in any drug regimen control strategy.

—Celina M. T. Martelli, Ms.C.
Ana Lucia S. S. Andrade, Ms.C.

*Departamento de Saude Coletiva
Instituto de Patologia Tropical
e Saude Publica
Universidade Federal de Goias, Brasil*

—Maria Aparecida F. Grossi, Ms.C.
Maria Ana A. Leboeuf, M.D.

*Secretaria Estadual de Saude
Minas Gerais, Brasil*

—Clovis Lombardi, Ph.D.
Fabio Zicker, Ph.D.

*Organizacion Panamericana
de la Salud, Venezuela*

Reprint requests to Dr. Celina Maria Turchi Martelli, Rua 132-A No. 74, Setor Sul, 74093-220 Goiania Goias, Brasil.

REFERENCES

1. CARTEL, J.L., SPIEGEL, A., NGOC, L.N., MOULIA-PELAT, J.P., MARTIN, P.M.V. and GROSSET, J.H. Leprosy in French Polynesia; the possible impact of multidrug therapy on epidemiological trends. *Lepr. Rev.* **63** (1992) 223-230.
2. FINE, P.E.M. Reflections on the elimination of leprosy. (Editorial) *Int. J. Lepr.* **60** (1992) 71-80.
3. IRGENS, L.M. and SKJAEVEN, R. Secular trends in age at onset, sex ratio, and type index in leprosy observed during declining incidence rates. *Am. J. Epidemiol.* **122** (1985) 695-705.
4. JESUDASAN, K., VIJAKUMARAN, P., PANNIKAR, V.K. and CHRISTIAN, M. Impact on leprosy as measured by selective indicators. *Lepr. Rev.* **59** (1988) 215-223.
5. LOMBARDI, C. Tendencia secular da detecção da hanseníase no estado de São Paulo. *Rev. Pat. Trop.* **22** (1993) 407-487.
6. LOMBARDI, C., COHEN, S., LEIKER, D.L., SOUZA, J.M.P., CUNHA, P.R., MARTELLI, C.M.T., ANDRADE, A.L.S.S. and ZICKER, F. Agreement between histopathological results in clinically diagnosed cases of indeterminate leprosy in São Paulo, Brazil. *Acta Leprol.* (1994). (in press)
7. MARTELLI, C.M.T., ANDRADE, A.L.S.S., SILVA, I.M., SOUZA, L., COSTA, M.B., MORAES NETO, O.L. and ZICKER, F. Considerações sobre classificação e definição de caso em hanseníase nos programas de controle. *Rev. Soc. Bras. Med. Trop.* **27** Suppl. 1 (1994) 21.
8. MARTELLI, C.M.T., MORAES NETO, O.L., ANDRADE, A.L.S.S., SILVA, S.A., SILVA, L.M. and ZICKER, F. Spatial patterns of Leprosy in an urban area in central Brazil. *Bull. WHO* (1994). (in press)
9. MOTTA, C.P. and ZUNIGA, M.G. Time trends of hansen's disease in Brazil. *Int. J. Lepr.* **58** (1990) 453-461.
10. NORDEEN, S.K. Leprosy 1962-992; Epidemiology and control of leprosy—a review of progress over last 30 years. *Trans. R. Soc. Trop. Med. Hyg.* **87** (1993) 515-517.
11. PANNIKAR, V.K. Defining a case of leprosy. *Lepr. Rev.* **63** Suppl. (1992) 61s-65s.
12. PATTYN, S.R. Future trends in the treatment of leprosy. *Trop. Geogr. Med.* **462** (1994) 85-88.
13. SMITH, P.G. Revised estimates of global leprosy numbers. *Lepr. Rev.* **62** (1992) 317-318.

1. CARTEL, J.L., SPIEGEL, A., NGOC, L.N., MOULIA-PELAT, J.P., MARTIN, P.M.V. and GROS-

Acid-Fast Bacilli from Former Leprosy Regions in Coastal Norway Showing PCR Positivity for *Mycobacterium leprae*

TO THE EDITOR:

Noncultivable acid-fast bacilli (NC-AFB) have been found in sphagnum samples collected in 1976 and 1977 in former leprosy

regions of coastal Norway⁽⁴⁾. Attempts to differentiate these NC-AFB have been made using white and nude mice and nine-banded armadillos. The results have indicated that

THE TABLE. Results of microscopic examination for acid-fast bacilli and PCR-based examination of sphagnum and musci samples from Norway.

| No. | Sphagnum or moss species | Microscopic examination ^a | PCR results |
|-----|--|--------------------------------------|-------------|
| 1. | <i>S. papillosum</i> | +ve | +ve |
| 2. | <i>S. magellanicum</i> | +ve | +ve |
| 3. | <i>S. magellanicum</i> | -ve | -ve |
| 4. | <i>S. papillosum</i> | -ve | -ve |
| 5. | <i>Drepanocladus</i> sp. | +ve | +ve |
| 6. | <i>S. magellanicum</i> | -ve | -ve |
| 7. | <i>Drepanocladus</i> sp. | +ve | +ve |
| 8. | <i>S. magellanicum</i> | +ve | +ve |
| 9. | <i>S. magellanicum</i> | -ve | -ve |
| 10. | <i>S. magellanicum</i> | +ve | +ve |
| 11. | <i>Rhytidiadelphus</i> sp. | -ve | -ve |
| 12. | <i>Pleurotium</i> (<i>Entodon</i>) <i>schreberi</i> | +ve | +ve |
| 13. | <i>S. subsecundum</i> | +ve | +ve |
| 14. | <i>S. rubellum</i> | -ve | -ve |

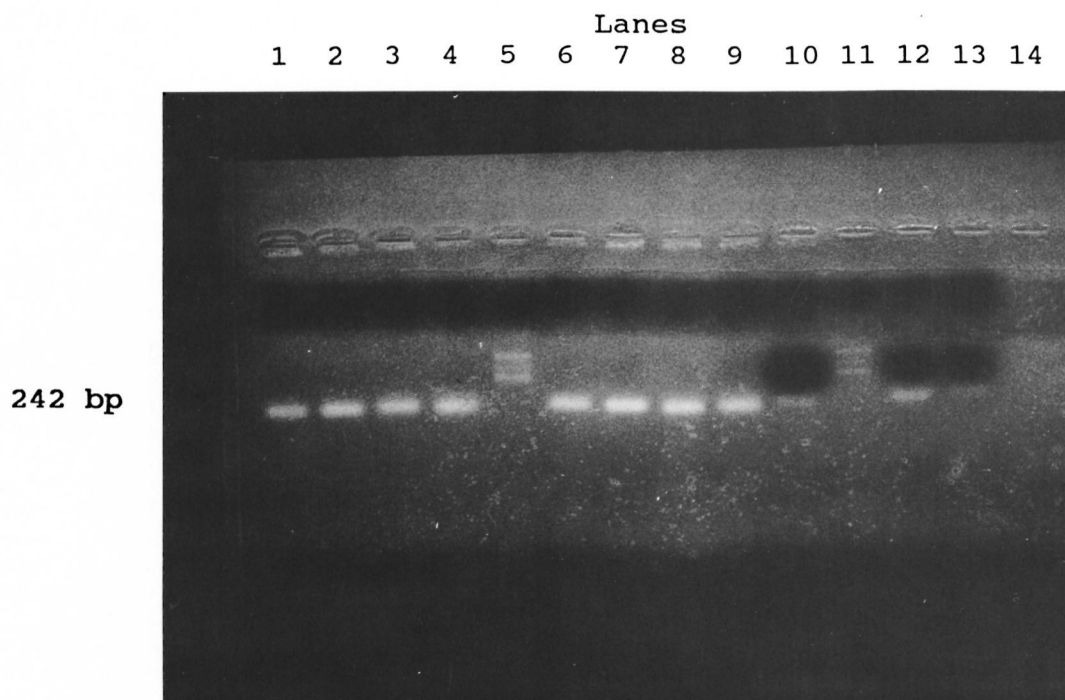
^a ≥ 3600 AFB/ml = +ve.

the isolated microorganism could not be distinguished from *Mycobacterium leprae* (1). In another study carried out in a leprosy

region of Bombay, *M. leprae* were found in polluted soil (2).

The second study in coastal Norway in 1986 succeeded in isolating NC-AFB with *M. leprae*-specific phenolic glycolipid-I in their cell wall. Experiments with nude mice indicated that these microorganisms had lost their pathogenicity (3). To clarify whether these microorganisms possess the same molecular genetic pattern as *M. leprae*, a third examination was carried out.

In October 1992, 14 samples of sphagnum and musci vegetation were collected on the island of Sotra near Bergen, Norway, in the same sphagnum bog where in both 1976 and 1986 NC-AFB were found (3, 4). The samples were collected with sterile gloves and kept in a refrigerator until processed. Sterile plastic syringes were filled with approximately 100 cc of the grey layer of the sample, compressed, fluid collected, and centrifuged at $8000 \times g \times 30$ min. To remove the humic acid and to reach a neutral reaction, the sediment was placed on a membrane filter, pore size $0.2 \mu\text{m}$, and washed 5 times with 100 ml of 1 M Tris



THE FIGURE. Positive reactions with *M. leprae*-specific primers in samples of sphagnum and musci vegetation. Lanes from the left: Samples nos. 1 (lane 1), 2 (lane 2), 5 (lane 3), 7 (lane 4), marker (lane 5), samples nos. 8 (lane 6) and 10 (lane 7). Positive controls: ML (lane 8), ML (lane 9), ML-dil. (lane 10), marker (lane 11), ML-dil. (lane 12), ML-dil. (lane 13). Negative control (lane 14). ML = armadillo-derived *M. leprae* 4.4×10^4 /ml; ML-dil. = armadillo-derived *M. leprae* 4.4×10^2 /ml.

buffer pH 8.5. The sediment was removed from the filter using 100 ml of 1 M Tris buffer pH 7.0 pressed in the opposite direction, the fluid centrifuged as above, and the sediment suspended in 0.5 ml of the last buffer solution. Smears were made, stained by the Ziehl-Neelsen (ZN) method for AFB, and the fluid used for further examination.

Lysis of the sample. In 1.5-ml screw-cap reaction cups (Eppendorf) containing 100 μ l of fluid, 50 μ l Proteinase K (in Tris buffer pH 8.5) and 50 μ l Tween 20 were added and covered with 50 μ l ultra-light mineral oil (Sigma) to prevent evaporation, and incubated in a water bath for 18 hr at 60°C. Thereafter, the samples were incubated at 97°C for 15 min to stop Proteinase K activity. The samples were frozen at -20°C until used.

Primers for polymerase chain reaction (PCR). The primers, found previously to be specific for *M. leprae* in a detailed comparison using *M. leprae* and 42 other mycobacterial species (5), were synthesized on an Applied Biosystems DNA synthesizer. The primers, SOD/I and SOD/II, were used to amplify a 242-base-pair fragment of the gene encoding for superoxide dismutase of *M. leprae*. The oligonucleotide primers have the following sequences: primer SOD/I (308) = 5'3'ATTGATGAAACGTTTGGGTC; primer SOD/II (530-550) = 3'5'TTGA-CGTAATCCGCCTTGAC.

Amplification of DNA by PCR. Twenty-five μ l of the samples to be analyzed was incubated in a microfuge tube containing 25 μ l of the reaction mixture [10 mM Tris-HCl (pH 9.6), 50 mM NaCl, 0.01% gelatin, 12.5 mM MgCl₂, 200 μ l of each of the oligonucleotide primers, 1 mM each of dATP, dCTP, dGTP, dTTP, and 2.5 units of Taq-polymerase]. Negative (with distilled water) and positive (with purified armadillo-derived *M. leprae*) samples were used as the controls. The reaction mixtures were covered with 25 μ l of ultra-light mineral oil (Sigma). Cycles of amplification consisted of a 2-min denaturation at 94°C, a 2-min annealing step at 59°C, and a 3-min elongation step at 72°C performed in a programmed DNA Thermal Cycler (automated Pharmacia LKB—Gene ATAQ Controller) and repeated 32 times. The samples were analyzed by electrophoresis (5).

Of 14 samples, 8 were ZN-positive and

the same samples revealed specific positivity for *M. leprae* (The Table). The Figure shows the results with 6 selected positive samples, armadillo-derived *M. leprae* in two concentrations (10⁴ and 10²) and one control. In the previous experiments, the low pH, caused by the presence of humic acid, hindered the action of Taq-polymerase (5). The modification of the procedure by washing with Tris buffer (pH 8.5) enabled the use of the PCR technique in sphagnum and musci samples.

The results confirmed that the NC-AFB present in sphagnum and musci vegetation possess the same fragment of the gene encoding for superoxide dismutase as *M. leprae*.

—Hamdy Mahfous Mostafa, M.D.

Faculty of Medicine
University of Assiut, Egypt

—Jindrich Kazda, M.V.D., Ph.D.

Professor
Institut für Experimentelle
Biologie und Medizin
D-23845 Borstel, Germany

—Lorentz M. Irgens, M.D.

Professor
Faculty of Medicine
University of Bergen, Norway

—Hans Gerd Luesse, Dr.Agr.

University Onsabruck, Germany

Reprint requests to Dr. Kazda.

REFERENCES

1. KAZDA, J. Occurrence of noncultivable acid-fast bacilli in the environment and their relationship to *M. leprae*. *Lepr. Rev.* **52** Suppl. 1 (1981) 85-91.
2. KAZDA, J., GANAPATI, R., REVANKAR, C., BUCHANAN, T. M., YOUNG, D. B. and IRGENS, L. M. Isolation of environment-derived *M. leprae* from soil in Bombay. *Lepr. Rev.* **57** Suppl. 3 (1986) 201-208.
3. KAZDA, J., IRGENS, L. M. and KOLK, A. Acid-fast bacilli found in sphagnum vegetation of coastal Norway containing *M. leprae*-specific phenolic glycolipid-I. *Int. J. Lepr.* **58** (1990) 353-357.
4. KAZDA, J., IRGENS, L. M. and MULLER, K. Isolation of noncultivable acid-fast bacilli in sphagnum and moss vegetation by foot pad technique in mice. *Int. J. Lepr.* **48** (1980) 1-6.
5. LUESSE, H. G. Untersuchungen zum Nachweis von *M. leprae* mit der Polymerase-Kettenreaktion. *Gottingener Beitr. Land-und Forstwirtschaft in den Tropen und Subtropen. Hefte* **78** (1993) 1-139.