Volume 69, Number 1 Printed in the U.S.A. (ISSN 0148-916X)

INTERNATIONAL JOURNAL OF LEPROSY and Other Mycobacterial Diseases

OFFICIAL ORGAN OF THE INTERNATIONAL LEPROSY ASSOCIATION

EDITORIAL OFFICE Gillis W. Long Hansen's Disease Center at Louisiana State University Baton Rouge, Louisiana 70894, U.S.A.

VOLUME 69, NUMBER 1

MARCH 2001

EDITORIAL

Editorial opinions expressed are those of the writers.

Factors Influencing the Development of Leprosy: An Overview

Leprosy is an infectious disease caused by an intracellular acid-fast bacterium: *Mycobacterium leprae*. In 1874, Armauer Hansen was the first to describe the bacterium as the cause of leprosy.³³ However, the triad of Koch is still not fulfilled. It has not been possible to infect someone willfully with *M. leprae*,⁵⁷ although anecdotal reports indicated infection after tattooing,⁷¹ dog bites, accidental inoculation^{68, 76, 78} and following the skinning and cleaning of infected armadillos for cooking.⁵⁵

CLINICAL SPECTRUM

There are various clinical manifestations of leprosy. However it is possible to classify the patients along a clinical spectrum. This was done elegantly coincidentally and independently by Ridley and Jopling⁷⁵ in the U.K. and by Leiker⁴⁸ in The Netherlands in 1966. The classification is based on the cell-mediated immune (CMI) response of the patients against *M. leprae*. At one end of the spectrum, the tuberculoid (TT) leprosy patients present with a relatively high CMI toward *M. leprae*, with one or a few well-defined hypopigmented or erythematous patches, usually with central healing and loss of sensation in the patch, and/or with an enlarged peripheral nerve. M. leprae are usually undetectable. At the other end of the spectrum, the lepromatous (LL) leprosy patients present with a complete tolerance to M. leprae and without any detectable CMI against the microbe. These patients are actually teeming with bacteria; they are the "perfect culture medium." The bacteria may be present anywhere in the body, with the possible exception of the central nervous system. The lepromatous patients may show ill-defined, minimal hypopigmented or erythematous patches, but sensation is still present. However they may show "glove-and-stocking" anesthesia with symmetrically enlarged peripheral nerves. They may also show nodules and plaques, skin colored or hyperpigmented, or show only a diffuse infiltration. There may be loss of eyebrows (madarosis) and a more-orless generalized diminished sweating. Between these two ends of the spectrum, the borderline leprosy group is found, encompassing most of the patients. The clinical range is from borderline tuberculoid (BT) leprosy with a few asymmetrically distributed, well-defined tuberculoid patches and a few enlarged nerves to borderline lepromatous (BL) leprosy with symmetrically distributed hypopigmented or erythematous macules and/or plaques, papules and nodules. The latter are mainly located on the cooler parts of the body. In the middle of the spectrum, mid-borderline (BB) leprosy patients have elevated lesions with an immune area (the center of the lesion is not involved) and typical dome-shaped, elevated small plaques.

In the borderline range, patients may upor downgrade (change their classification within the spectrum). Upgrading indicates that the patient develops more tuberculoid features; downgrading, more lepromatous. In upgrading leprosy the bacterial load diminishes; in downgrading the bacterial load is increased by bacterial multiplication. In a downgraded patient, a few of the older patches may show loss of sensation; whereas the new lesions do not. In an upgrading patient, new tuberculoid-like lesions may appear or the lesions may become atrophic (heal).

Upgrading and downgrading occurs either silently or is accompanied by a reactional phenomenon called reversal reaction (RR), in which an enhanced CMI toward *M. leprae* antigenic determinants may cause irreversible nerve damage.⁶⁴

Indeterminate leprosy comprises a special group of leprosy patients having one or two slightly hypopigmented or erythematous macules with or without detectable loss of sensation or loss of sweating. The biopsy may show a single bacterium or a minimal lymphocytic infiltration in a dermal nerve. The diagnosis is difficult to establish, and some leprologists consider it to be an early form of either multibacillary (MB) or paucibacillary (PB) leprosy which may either heal (over 80%) or become frank MB or PB leprosy.⁶⁶

MODE OF INFECTION AND DEVELOPMENT OF DISEASE

The mode of infection is still a point of discussion. Most leprologists no longer consider the skin to be important as the port of entry or exit of *M. leprae*.³⁹ However, it was recently reported again that a marked number of *M. leprae* is present in all layers

of the epidermis, including the stratum corneum in lepromatous leprosy patients.⁴⁰ It may well be that this "exit" has been neglected since the reports by Pedley on the nonemergence of M. leprae from intact lepromatous skin⁷⁰ and later by Rees and Meade on the possibility of airborne infections.73 Nonetheless, some leprologists and pathologists still continue to consider it to be a real possibility.⁴⁹ As port d'entrée, the skin is only mentioned in anecdotal reports of infection occurring after tattooing,⁷¹ dog bites and accidental inoculation^{68, 76, 78} or after the skinning of infected armadillos.55 There are also numerous observations of a first patch on the forehead or on the cheek of a baby carried on the back of its lepromatous mother, and the first lesions seen on the bare buttocks of toddlers sitting on contaminated soil. Horton and Povey35 concluded that the distribution of the first lesion is not at random but confined to exposed parts of the body. This concept was recently supported by Abraham, et al.² who concluded that the first lesions occur exactly at the sites most vulnerable to trauma. Naafs⁶¹ showed for Ethiopia that the age at onset of leprosy between 1973-1979 followed the same pattern as that of tetanus, excluding neonatal tetanus, when allowing an incubation period of between 2-5 years for leprosy. It also has been shown that contaminated thorns may infect susceptible mice.38

Insect bites have long been incriminated in the transmission of leprosy.^{36, 76, 87, 88} However in experimental studies it proved ineffective though possible.^{5, 65, 68, 89} As a possible route of infection it cannot be fully dismissed. Moreover vomits of insects which had ingested *M. leprae* were shown to contain acid-fast material.⁷⁶ Flies were able to transport *M. leprae* on their feet.³⁰

Transmission via the gastrointestinal tract received some attention because *M. leprae* was found to be present in mothers' milk.^{69,82} However, epidemiological evidence for this route of infection is lacking.²⁵ In an experimental set up neither in Carville¹⁹ nor in London^{46, 54, 68} could this route of infection be proven, although viable bacilli were seen in the stool of the challenged animals. Sexual transmission has often been considered,⁷⁸ but being a complex contact, the route is not clear. However the vaginal mucosa of lepromatous women and the penile head of lepromatous men showed numerous acid-fast mycobacteria.⁵⁹

Leprosy is at present considered to be an airborne disease having a transmission pattern similar to that of tuberculosis, in which infectious patients or carriers discharge bacteria from the nasal mucosa.6 Rees and Meade elegantly showed this possibility.73 Some authors were doubtful because the age at onset in their particular environment was significantly earlier for leprosy than for tuberculosis, although both diseases were highly endemic.⁶⁰ As port d'entrée, the respiratory tract has been suggested, with the nose playing a central role. Rees and Mc-Dougall⁷⁴ showed such port d'entrée to be possible for thymectomized mice; Chehl, et al.¹⁹ for nude mice and, more recently, Vilani-Moreno, et al.98 confirmed this for the immune-competent Swiss mice. The central role of the nose may be illustrated by the observation by Cerotti¹⁶ that only 14 out of 116 mucosal biopsies showed to be normal¹⁶ and that even in "pure neural leprosy' more than half of the patients show inflammatory changes in their nasal mucosa.94

It still remains unknown, however, why certain individuals develop leprosy and others do not. For a long time leprosy was considered to be an inherited disease,11 until Armauer Hansen showed it to be an infectious one.33 However, the observation that leprosy often affects families,³⁴ which cannot always be explained by a more intensive exposure, still holds. Rotberg proposed a theoretical, inherited, N-factor.79,80 Beiguelman showed a family association of Mitsuda positivity.7-10 Of interest in this respect is the observation that the Nramp1 homolog seems to be associated with a granulomatous Mitsuda reaction.³ That it could not be a simple straight forward inherited factor like, for instance, the factor that codes for epidermodysplasia verruciformis was shown in twin studies.18

An innate immunity has been proposed for some of the infected individuals.⁹⁰ For the majority, however, the CMI seems to be of crucial importance. For a short period of time, it was thought that the HLA-DR loci were the decisive factors,⁹⁹ but this was soon challenged.⁹² Later, it was shown that both HLA-DR phenotypes 1^{13, 56} and 2 had some influence on the type of leprosy that develops after infection, but had no influence on whether or not someone developed leprosy.^{13, 14, 22, 23, 67, 100} Feitosa, et al.,²⁴ using complex segregation analyses of 10,886 individuals distributed among 1568 families, concluded that there might be a recessive major gene controlling susceptibility. However, they could not find evidence for unique genetic determinants for the leprosy subtypes, although they found indications of a segregating major effect between tuberculoid and lepromatous. Recently Silva, et al.⁸⁶ investigated the Lewis blood group phenotypes in leprosy patients and showed that nonsecretors developed significantly more leprosy than secretors. This finding suggests that the glycoprotein that is coded for, when secreted in the nasal mucous, has a protective action, possibly hindering adherence of M. leprae to the mucosal surface by binding to the adherence sites on the bacterium. A similar possibility can also be proposed for urinary tract infections,83 recurrent vulvovaginal candidiasis17 and pyloribacterium infections which lead to gastric ulcers.41

A polymorphism in a nucleotide relative to the transcriptional start site of tumor necrosis factor (TNF), a critical mediator of host defense and pathology, has been associated with lepromatous leprosy, as well as with severe malaria, leishmaniasis and scarring glaucoma.44, 81 Subtle mutations in pathways leading to cytokine or chemokine production or receptor presentation also have been suggested as possible mechanisms that could play a role in susceptibility to infections such as tuberculosis and leprosy.50 The same applies for factors involved in the milieu interior of cells. Allelic variants which seem to be related to innate immunity, at the human Nramp1 homolog, have recently been found to be associated with susceptibility to these two infections.3,12

Mucosal, secretory IgA, immunity is another factor that could influence the protection against, or the maintenance of, intranasal infection.^{1, 21, 72} It was found that workers at a leprosy hospital had a high level of secretory IgA against *M. leprae*; whereas lepromatous leprosy patients did not.²⁰ An interesting finding is that secretory IgA secretion is enhanced by stimulation of both sympathetic and parasympathetic nerves.¹⁵ Nerves are noted to be damaged throughout the leprosy spectrum, but most of all in lepromatous leprosy patients.²⁹

It has been suggested that the port of entry of *M. leprae* antigenic determinants may be important for the immune sys-tem,^{25, 45, 61, 62, 68, 84, 93} as supported by a concept assuming a peripheral and a central lymphocyte compartment.93 An encounter via the skin and the draining lymph nodes (peripheral compartment) stimulates CMI. A stimulus via the nerve directly into the peripheral blood/spleen (central compartment) leads to an immunosuppression, and may induce tolerance.93 More recently, it has been shown that exposure to antigens in the nasal mucosa also can lead to an immune tolerance.^{32, 85, 95, 96} This is even more interesting when one realizes that in an endemic community 5% or even up to 27% of the population may harbor M. leprae in their nose, as shown in a polymerase chain reaction (PCR) for *M. leprae* DNA.^{37, 43} Even some visitors from nonendemic countries who worked for a period of time in a leprosy hospital have been shown to have transient positive nose swabs for M. leprae DNA.63 A factor in this may be the Lewis phenotype⁸⁶ hindering or facilitating adherence to the nasal mucosa and the presence or absence of anti-M. leprae secretory IgA.21

It has been established that M. leprae are able to survive for several weeks (2-4) in the environment, especially under moist conditions.42 Such conditions exist in and around living quarters in many of the endemic countries.76 In most of these countries, blowing one nostril while closing the other cleans noses. The mucus will partly disperse, but most of it together with M. leprae reaches the ground. Contaminated epidermal corneal scales may also accumulate here. Kazda, et al.,42 using the mouse foot pad culture, showed the presence of M. leprae in soil. Matsuoka, et al.53 found M. leprae DNA in nearly half of the water samples tested in a leprosy-endemic area. There was a higher prevalence of leprosy among the people that used this water for bathing and washing. Toddlers sit, crawl and play on and in these contaminated environments, sustaining small injuries.

Children are prone to itch because they are in the process of immunological adaptation to their physical environment. They easily scratch themselves after contact with insects and other parasites, thereby introducing *M. leprae* from the soil or other sources with their nails into their skin. This inoculation into a part of the peripheral lymphocyte compartment may stimulate CMI. Acid-fast material (possibly bacteria) was found under the nails of children,59 whether it was M. leprae could not be established at the time. The contact with M. leprae-shedding family members or visitors also may be of a more direct nature. They may discharge M. leprae in large amounts in an aerosol³¹ as already shown by Schaeffer early in the 20th century.97 The bacterium may then enter the nasal mucosa of a child and induce tolerance. The observation of Fokkens, et al.29 that leprosy patients have a diminished number of CD8+ cytotoxic T cells in their nasal mucosa may be important. Whether this is the consequence of the infection, a facilitating factor or both could not be established. It was also noticed that the mucosa was atrophic and damaged with blood vessels very near to the surface,²⁹ thus providing easy access for the bacterium to the central lymphocyte compartment. It should be realized that not only the route of the infection but also the size, the viability, the interval and the frequency of the inoculum are important.45 Little is known on this subject to date.

Not only *M. leprae* but also environmental mycobacteria may have an influence on the immune system.^{4, 47, 52, 58} Auto-antigens, too, may modify the immune response. The influence of BCG vaccination is well known,^{26–28, 51, 77} its effectiveness probably depending on the environmental microorganisms.^{27, 28, 91}

CONCLUSION

It may be theorized that the balance between responses elicited by different routes of infection and inoculum, skin versus nasal mucosa and possibly nerve,⁹³ is responsible for the outcome of the infection. However, data to date suggest that the response is modulated by genetic factors, among which is HLA-DR. Even more important are previous encounters with other microorganisms and auto-antigens with antigenic determinants similar to those of *M. leprae*. The final result, resistance, delayed-type hypersensitivity, tolerance, disease or no disease, tuberculoid, borderline or lepromatous leprosy with or without reactions, is most likely mediated by the orchestration of the induced cyto- and chemokines.⁶⁴

SUMMARY

The clinical manifestations of leprosy vary, seemingly depending on the host's immune response. Mode and route of infection, such as skin versus nasal mucosa, insect bites, sexual and gastroenteral transmission, together with genetic factors that may contribute to the outcome of the infection, including HLA, Lewis factor, Nramp1 and more subtle inherited alterations, are discussed. It is theorized that a balance between host responses elicited by different routes of infection and size and spacing of inocula is responsible for the clinical and immunological manifestations of the disease. Genetic factors and contact with environmental microorganisms may modulate these responses. The final result, resistance, delayed-type hypersensitivity, tolerance, disease or no disease, spectrum and reactions, is most likely reached via the orchestration of the induced cyto- and chemokines.

-Ben Naafs, M.D., Ph.D.

Instituto Lauro de Souza Lima Bauru, SP, Brazil and Department of Dermatology Leiden University Medical Center Leiden, The Netherlands

> —Eliane Silva, Fatima Vilani-Moreno, Elaine Camarinha Marcos, Maria Esther Nogueira, Diltor V. A. Opromolla, M.D., Ph.D.

Instituto Lauro de Souza Lima Bauru, Sao Paulo, Brazil

Reprint requests to B. Naafs, M.D., Ph.D., Dermato-venereologist, Gracht 15, 8485 KN Munnekeburen, The Netherlands. Tel/Fax: 31-561-481595; e-mail: benaafs@dds.nl

Acknowledgment. This contribution was made possible by the unpaid leave granted to Dr. B. Naafs by the IJsselmeerziekenhuizen. The QM Gastmann Wichersstichting is thanked for paying the airfare to Brazil. Dr. B. Tank advised on the use of the English language.

REFERENCES

- ABE, M., YOSHINO, Y., MINAGAWA, F., MIYAJI, L., SAMPOONACHOT, P., OZAWA, T., SAKAMOTO, Y., SAITO, T. and SAIKAWA, K. Salivary immunoglobulins and antibody activities in leprosy. Int. J. Lepr. 52 (1984) 343–350.
- ABRAHAM, S., MOZHI, N. M., JOSEPH, G. A., KURIAN, N., SUNDAR RAO, S. S. and JOB, C. K. Epidemiological significance of first skin lesion in leprosy Int.J. Lepr. 66 (1998) 131–139.
- ALCAIS, A., SANCHEZ, F. O., THUC, N. V., LAP, V. D., OBERTI, J., LAGRANGE, P. H., SCHURR, E. and ABEL, L. Granulomatous reaction to intradermal injection of lepromin is linked to the human NRAMP1 gene in Vietnamese siblings. J. Infect. Dis. **182** (2000) 302–308.
- ARANDA, C. M., CHIN-A-LIEN, R. A. M., KANT, M. J., KOLK, A. J. H., OPROMOLLA, D. V. A., TANK, B. and NAAFS, B. Environmental mycobacteria may induce recognition of auto-antigens: circumstantial evidence using maternal and cord blood (submitted for publication).
- BANERJEE, R., BANERJEE, B. D., CHAUDHURY, S. and HATI, A. K. Transmission of viable *Mycobacterium leprae* by *Aedes aegypti* from lepromatous leprosy patients to the skin of mice through interrupted feeding. Lepr. Rev. 62 (1994) 21–26.
- BARTON, R. P. E. A clinical study of the nose in leprosy. Lepr. Rev. 45 (1974) 135–144.
- 7. BEIGUELMAN, B. and QUAGLIATO, R. Nature and familial character of the lepromin reactions. Int. J. Lepr. **33** (1965) 800–807.
- BEIGUELMAN, B. The genetics of resistance to leprosy. Int. J. Lepr. 33 (1965) 808–812.
- BEIGUELMAN, B. Lepromin reaction: genetic studies including twin pair analysis. Acta Leprol. 44 (1971) 5–65.
- BEIGUELMAN, B. A reação de Mitsuda oitenta anos depois. Hansen. Int. 24 (1999) 144–161.
- 11. BOECK, C. W. and DANIELSSEN, D. C. Om spedalskhed. Christiana, Norway, 1847.
- CANONNE-HERGAUX, F., GRUENHEID, S., GOVONI, G. and GROS, P. The Nramp1 protein and its role in resistance to infection and macrophage function. Proc. Assoc. Am. Physicians 111 (1999) 283–289.
- CAMARINHA MARCOS, E.V. Imunogenetica In: Noções de Hansenologia. Opromolla, D.V.A., ed. Bauru, Brazil: ILSL, 2000 pp. 43–46.
- CAMARINHA MARCOS, E. V., SOUZA, F. C., URA, S. and OPROMOLLA, D. V. A. Estudo de associaço entre antigenos HLA e reacço hansenica tipo 1 ulcerada. An. Bras. Dermatol., Rio de Janeiro 75 (2000) 283–290.
- CARPENTER, G. H., GARRETT, J. R., HARTLEY, R. H. and PROCTOR, G.B. The influence of nerves on the secretion of immunoglobulin A into submandibular saliva in rats. J. Physiol. (Lond.) **512** (1998) 567–573.
- CERRUTI, H. Histopathology of the nasal mucosa in leprosy. Rev. Bras. Leprol. 12 (1944) 309–364.

- CHAIM, W., FOXMAN, B. and SOBEL, J. D. Association of recurrent vaginal candidiasis and secretory ABO and Lewis phenotype. J. Infect. Dis. **176** (1997) 828–830.
- CHAKRAVARTTI, M. R. and VOGEL, F. A twin study on leprosy. In: *Topics in Human Genetics, Vol. 1.* Becker, P. E., Lenz, W., Vogel, F. and Wendt, G., eds. Stuttgart: Thieme, 1973.
- CHEHL, S., JOB, C. K. and HASTINGS, R. C. Transmission of leprosy in nude mice. Am. J. Trop. Med. Hyg. 34 (1985) 1161–1166.
- CREE, I. A., SHARPE, S., STURROCK, N. D. C., COCHRANE, I. H., SMITH, W. C. S. and BECK, J. S. Mucosal immunity to mycobacteria in leprosy patients and their contacts. Lepr. Rev. 59 (1988) 309–316.
- CREE, I. A. and SMITH, W. C. Leprosy transmission and mucosal immunity: towards eradication? Lepr. Rev. 69 (1998) 112–121.
- 22. VAN EDEN, W., DE VRIES, R. R., MEHRA, N. K., VAIDYA, M. C., D'AMARO, J. and VAN ROOD, J. J. HLA segregation of tuberculoid leprosy: conformation of the DR2 marker. J. Infect. Dis. 141 (1980) 693–701.
- VAN EDEN, W., GONZALEZ, N. M., DE VRIES, R. R., CONVIT, J. and VAN ROOD, J. J. HLA-linked control of predisposition to lepromatous leprosy. J. Infect. Dis. 151 (1985) 9–14.
- FEITOSA, M. F., BORECKI, I., KRIEGER, H., BEIGUELMAN, B. and RAO, D. C. The genetic epidemiology of leprosy in a Brazilian population. Am. J. Hum. Genet. 56 (1995) 1179–1185.
- FINE, P. E. M. Leprosy: the epidemiology of a slow bacterium. Epidemiol. Rev. 4 (1982) 161–188.
- FINE, P. E. M., STERNE, J. A., PONNINGHAUS, J. M. and REES, R. J. Delayed-type hypersensitivity, mycobacterial vaccines and protective immunity. Lancet 344 (1994) 1245–1249.
- FINE, P. E. M. and TRUMAN, R. Report of workshop on epidemiology/transmission/vaccines. Int. J. Lepr. 66 (1998) 596–597.
- FINE, P. E. M. and VYNNYCKY, E. The effect of heterologous immunity upon the apparent efficacy of (e.g. BCG) vaccines. Vaccine 16 (1998) 1923–1928.
- FOKKENS, W. J., NOLST TRINITE, G. J., VIRMOND, M., KLEINJAN, A., ANDRADE, V. L. G., VAN BAAR, N. G. and NAAFS, B. The nose in leprosy; immunohistopatholology of the nasal mucosa. Int J. Lepr. 66 (1998) 328–339.
- GEATER, J. G. The fly as a potential vector in the transmission of leprosy. Lepr. Rev. 46 (1975) 279–286.
- GREEN, C. A., KATOCH, V. M. and DESIKAN, K. V. Quantitative estimation of *Mycobacterium leprae* in exhaled nasal breath. Lepr. Rev. 54 (1983) 337–340.
- HANNINEN, A. and HARRISON, L. C. Gamma delta T cells as mediators of mucosal tolerance: the autoimmune diabetes model. Immunol. Rev. 173 (2000) 109–119.

- HANSEN A. G. Spedalskhedens aarsager. Norsk Mag. f. Leagavid 4 (1874) 76–79.
- 34. HOPKINS, R. and DENNY, O. E. Leprosy in the United States. J. Amer. Med. Assoc. **92** (1929) 191–192.
- HORTON, R. J. and POVEY, S. The distribution of the first lesion in leprosy. Lepr. Rev. 37 (1966) 113–114.
- 36. 2TE INTERNATIONALE WISSENSCHAFTLICHEN LEPRA-KONFERENZ IN BERGEN 1909. Mitteilungen und Verhandlungen von Dr H.P.Lie. Band III Leipzig: J.A. Barth, 1910, p. 416.
- IZUMI, S., BUDIAWAN, T., SAEKI, K., MATSUOKA, M. and KAWATSU, K. An epidemiological study on *Mycobacterium leprae* infection and prevalence of leprosy in endemic villages by molecular biological technique. Indian J. Lepr. **71** (1999) 37–41.
- JOB, C. K., CHEHL, S. K. and HASTINGS, R. C. Transmission of leprosy in nude mice through thorn pricks. Int. J. Lepr. 62 (1994) 395–398.
- 39. JOB, C. K., BASKARAN, B., JAYAKUMAR, J. and ASCHOFF, M. Histopathologic evidence to show that indeterminate leprosy may be a primary lesion of the disease. Int. J. Lepr. 65 (1997) 443–450.
- JOB, C. K., JAYAKUMAR, J. and ASCHOFF, M. "Large numbers" of *Mycobacterium leprae* are discharged from the intact skin of lepromatous patients; a preliminary report. Int. J. Lepr. 67 (1999) 164–167.
- KABAYASHI, C., SAKAMOTO, J., KITO, T., YAMA-MURA, Y., KASHIKAVA, T., FUJITA, M., WATANABE, T. and NAKAZATO, H. Lewis blood group-related antigen expression in normal gastric mucosa, intestinal metaplasia, gastric adenoma and gastric carcinoma. Am. J. Gastroenterol. 88 (1993) 919–924.
- KAZDA, J., GANAPATI, R., REVANKAR, C., BUCHAN-AN, T. M., YOUNG, D. B. and IRGENS, L. M. Isolation of environment-derived *Mycobacterium leprae* from soil in Bombay. Lepr. Rev. 57 (1986) 201–208.
- KLATSER, P. R., VAN BEERS, S. M., MADJID, B., DAY, R. and DE WIT, M. Y. L. Detection of mycobacterial carriage in a leprosy endemic population, J. Clin. Microbiol. 31 (1993) 2947–2951.
- KNIGHT, J. C. and KWIATKOWSKI, D. Inherited variability of tumor necrosis factor production and susceptibility to infectious disease. Proc. Assoc. Am. Physicians 111 (1999) 290–298.
- LAGRANGE, P. H., MACKANESS, G. B. and MILLER, T. E. Influence of dose and route of antigen injection in the immunological induction of T-cells. J. Exp. Med. **193** (1974) 528–542.
- 46. LANCASTER, R. P. Development and use of the nude mouse as a model of lepromatous leprosy. Ph.D. Thesis, University of London, U.K., 1985.
- 47. DE LANGE, W. E., GWANZURA, L., MOUS, H. V. M., MASON, P. R. and NAAFS, B. Sensitisation to mycobacteria in two areas of Zimbabwe with different distribution of leprosy clinical type and lep-

69, 1

rosy incidence: ELISA. (Congress Abstract) Int J. Lepr. **61** (1993) 100A.

- LEIKER, D. L. Classification of leprosy. Lepr. Rev. 38 (1966) 7–15.
- LEIKER, D. L. On the mode of transmission of *My*cobacterium leprae. Lepr. Rev. 48 (1977) 9–16.
- LEVIN, M. and NEWPORT, M. Understanding of the genetic basis of susceptibility to mycobacterial infection. Proc. Assoc. Am. Physicians. 111 (1999) 308–312.
- LOMBARDI, C., PEDAZZANI, E. S., PEDRAZZANI, J. C., FILHO, P. F. and ZICKER, F. Protective efficacy of BCG against leprosy in Sao Paulo. Bull. Pan. Am. Health Organ. **30** (1996) 24–30.
- LYONS, N. F. and NAAFS, B. Influence of environmental mycobacteria on the prevalence of leprosy clinical type. Int. J. Lepr. 55 (1987) 637–645.
- 53. MATSUOKA, M., IZUMI, S., BUDIAWAN, T., NAKATA, N. and SAEKI, K. *Mycobacterium leprae* DNA in daily using water as a possible source of leprosy infection. Indian J. Lepr. **71** (1999) 61–74.
- MCDERMOTT-LANCASTER, R. D. and MCDOUGALL, A. C. Mode of transmission and histology of *M. leprae* infection in nude mice. Int. J. Exp. Pathol. 71 (1990) 689–670.
- 55. MEYERS, W. M. Leprosy. Dermatol. Clin. 10 (1992) 73–96.
- MIYANAGA, K., JUJI, T., MAEDA, H., NAKIJIMA, S. and KOBAYASHI, S. Tuberculoid leprosy and HLA in Japanese. Tissue Antigens 3 (1981) 331–334.
- MORETSZOHN, L. B. L. *Etiologia e pathogenia da lepra*. M.D. thesis, Faculdade Medicina do Rio de Janeiro, Brazil, 1907.
- MOUS, H. V. H., MASON, P. R., DE LANGE, W. E., GWANZURA, L. and NAAES, B. Sensitisation to mycobacteria in two areas in Zimbabwe with different distribution of leprosy type and leprosy incidence: skin tests. (Congress Abstract) Int. J. Lepr. 61 (1993) 100A.
- NAAFS, B. Unpublished observations, ALERT, Addis Ababa, Ethiopia, 1978.
- NAAFS, B. and ADEMU ALEMAYU. Comparison of the modes of spread and the age of onset of leprosy and tuberculosis. In: *Nerve Damage in Leprosy*. Naafs, B. Ph.D. thesis, University of Amsterdam, The Netherlands, 1980, VI 20–VI 25.
- NAAFS, B. Leprosy: general introduction. In: *Nerve Damage in Leprosy.* Naafs, B. Ph.D. thesis, University of Amsterdam, The Netherlands, 1980, II 2–II 27.
- NAAFS, B. Eziopatogenesi. In: *Manuale di Leprologia*. Nunzi, E. and Leiker, D.L., eds. Bologna: OCSI, 1990, pp. 47–50.
- NAAFS, B., KLATSER, P. R. VIRMOND, M. and FOKKENS, W. J. Unpublished observations, Instituto Lauro de Souza Lima, Bauru, Brazil, 1997.
- NAAFS, B. Current views on reactions in leprosy. Indian J. Lepr. 72 (2000) 97–122.
- 65. NARAYANAN, E., SREEVATSA, KIRCHHEIMER, W. F. and BEDI, B. M. S. Transfer of leprosy bacilli from

patients to mouse foot pads by *Aedes aegypti*. Lepr. India **49** (1977) 181–186.

- OPROMOLLA, D. V. A. Classificacao. In: *Noções de Hansenologia*. Opromolla, D.V.A., ed. Bauru, Brazil: ILSL, 2000, pp. 47–51.
- 67. OTTENHOFF, T. M., GONZALEZ, N. M., DE VRIES, R. R. P., CONVIT, J. and VAN ROOD, J. J. Association of HLA specificity LB-E12 (MB1, DC1, MT1) with lepromatous leprosy in a Venezuelan population. Tissue Antigens 24 (1984) 25–29.
- PALLEN, M. J. and MCDERMOTT, R. D. How might *Mycobacterium leprae* enter the body. Lepr. Rev. 57 (1986) 289–297.
- 69. PEDLEY, J. C. The presence of *M. leprae* in human milk. Lepr. Rev. **38** (1967) 239–242.
- PEDLEY, J. C. Composite skin contact smears: a method of demonstrating the non-emergence of *Mycobacterium leprae* from intact lepromatous skin. Lepr. Rev. 41 (1970) 31–43.
- PORRITT, R. J. and OSLEN, R. S. Two simultaneous cases of leprosy developing in tattoos. Am. J. Pathol. 23 (1947) 805–817.
- 72. RAMAPRASAD, P., FERNANDO, A., MADHALE, S., RAO, J. R., EDWARD, V. K., SAMSON, P. D., KLATSER, P. R., DE WIT, M. Y. L., SMITH, W. C. S. and CREE, I. A. Transmission and protection in leprosy; indications of the role of mucosal immunity, Lepr. Rev. 68 (1997) 301–315.
- REES, R. J. W. and MEADE, T. W. Comparison of the modes of spread and the incidence of tuberculosis and leprosy. Lancet 1 (1974) 47–49.
- REES, R. J. W. and MCDOUGALL, A. C. Airborne infection with *Mycobacterium leprae* in mice. J. Med. Microbiol. **10** (1977) 63–68.
- 75 RIDLEY, D. S. and JOPLING, W. H. Classification of leprosy according to immunity; a five-group system. Int. J. Lepr. 34 (1966) 255–273.
- RIVAS, G. M. Estudios Experimentales Sobre la Transmision de la Lepre. Bogota: Lab. Munoz-Rivas, 1958.
- 77. RODRIGUES, M. L. O., SILVA, S. A., NETO, J. C., DE ANDRADE, A. L., MARTELLI, C. M. and ZICKER, F. Protective effect of intradermal BCG against leprosy; a case-control study in central Brazil. Int. J. Lepr. **60** (1992) 335–339.
- ROGERS, L. and MUIR, E. Contagiousness of leprosy. In: *Leprosy.* 2nd edn. Bristol: John Wright, 1940, pp. 71–79.
- ROTBERG, A. Some aspects of immunity in leprosy and their importance in epidemiology, pathogenesis and classification of forms of the disease. Rev. Bras. Leprol. 5 (1937) 45–97.
- ROTBERG, A. Fiftieth anniversary of the "N-factor/Hansen-anergic fringe" hypothesis for hanseniasis. Int. J. Lepr. 57 (1989) 864–866.
- ROY, S., MCGUIRE, W., MASCIE-TAYLOR, C. G., SAHA, B., HAZRA, S. K., HILL, A. V. and KWIAT-KOWSKI, D. Tumor necrosis factor promotor polymorphism and susceptibility to lepromatous leprosy. J. Infect. Dis. **176** (1997) 530–532.

Editorial

- SAHA, K., SHARMA, V. and SIDDIQUI, M. A. Decreased cellular and humoral anti-infective factors in the breast secretions of lactating mothers with lepromatous leprosy. Lepr. Rev. 53 (1982) 35–44.
- SHEINFIELD, J., SCHAEFFER, A. J., CORDONCARDO, C., ROGATKO, A. and FAIR, W. R. Association of the Lewis-blood phenotype with recurrent urinary tract infections in women. N. Engl. J. Med. 320 (1989) 773–777.
- SHEPARD, C. C., WALKER, L. L., VAN LANDING-HAM, R. M. and YE, S. Z. Sensitisation or tolerance to *Mycobacterium leprae* antigens by route of injection. Infect. Immuun. **38** (1982) 673–680.
 SHI, F. D., LI, H., WANG, H., BAI, X., VAN DER MEIDE, P. H., LINK, H. and LJUNGGREN, H. G. Mechanisms of nasal tolerance induction in experimental autoimmune myasthenia gravis; identification of regulatory cells. J. Immunol. **162** (1999) 5757–5763.
- SILVA, E., RUBIO, E. M. and URA, S. Sistema sanguineo Lewis em patientes hansenianos. Hansen. Int. (in press 2001).
- SOUZA AROUJO, H. C. Infecção espontânea e experimental de hematofagos em leprosos. Mem. Inst. Oswaldo Cruz 38 (1943) 447–463.
- SOUZA AROUJO, H. C. Da lepra; sua provavel transmissio pelos artropodos. Arq. Minais. Leprol. 13 (1953) 42–58.
- SREEVATSA. Leprosy and arthropods. Indian J. Lepr. 65 (1993) 189–200.
- STACH, J. L., DELGADO, G., TSCHIBOZO, V., STRO-BEL, M. and LAGRANGE, P. H. Natural resistance to mycobacteria: antimicrobial activity and reactive oxygen intermediate releasing functions of murine macrophages. Ann Immunol. (Paris) 135 (1984) 25–37.
- STANFORD, J. L., SHIELD, M. J. and ROOK, G. A. How environmental mycobacteria may predetermine the protective influence of BCG. Tubercle 62 (1981) 55–62.

- 92. STONER, G. L., TOUW J., BELEHU, A. and NAAFS, B. *In-vitro* lymphoproliferative response to *My-cobacterium leprae* of HLA-D-identical siblings of lepromatous leprosy patients. Lancet **8089** (1978) 543–547.
- STONER, G. L. Importance of the neural predilection of *Mycobacterium leprae* in leprosy. Lancet 8150 (1979) 994–996.
- SUNEETHA, S., ARUNTHATHI, S., JOB, A., DATE, A., KURIAN, N. and CHACKO, C. J. G. Histological studies in primairy neuritic leprosy: changes in the nasal mucosa. Lepr. Rev. 69 (1998) 358–366.
- 95. TIAN, J., ATKINSON, M. A., CLARE-SALZLER, M., HERSCHENFELD, A., FORSTHUBER, T., LEHMANN, P. V. and KAUFMAN, D. L. Nasal administration of glutamate decarboxylase (GAD65) peptides induces Th2 responses and prevents murine insulin-dependant diabetes. J. Exp. Med. 183 (1996) 1561–1567.
- TSITURA, D. C., DEKRUYFF, R. H., LAMB, J. R. and UMETSU, D. T. Intranasal exposure to protein antigens induces immunological tolerance mediated by functionally disabled CD4+ T cells. J. Immunol. 163 (1999) 2592–2600.
- URA, S. and OPROMOLLA, D. V. A. Epidemiologia. In: *Noções de Hansenologia*. Opromolla, D.V.A., ed. Bauru, Brazil: ILSL, 2000, pp. 109–112.
- VILANI-MORENO, F. R., ARRUDA, M. S. P., NOGUEIRA, M. E. S. and BAPTISTA, I. M. F. D. Hanseniase experimental murina: inoculação do *Mycobacterium leprae* via intranasal. Hansen. Int. 24 (1999) 115–120.
- 99. DE VRIES, R. R. P., LAI-A-FAT, R. F., NIJENHUIS, L. E. and VAN ROOD, J. J. HLA-linked genetic control of host response to *Mycobacterium leprae*. Lancet **7999** (1976) 1328–1330.
- 100. DE VRIES, R. R. P., MEHRA, N. K., VAIDYA, M. C., GUPTE, M. D., KAHN, P. M. and van Rood, J. J. HLA-linked control of susceptibility to tuberculoid leprosy and association with HLA-DR types. Tissue Antigens 16 (1980) 294–304.